

Antimutagenic effect of grape seed extracted oil on diazinon induced genotoxicity in albino mice

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Abstract:

Background: Human health hazard due to exposure to chemical pollutants is a global and chronic problem. The pesticides constitute the primary toxic chemicals in the environment. The present study explored the effects of grape seed oil supplementation on diazinon (DZN) induced chromosomal aberrations and sperm abnormalities in male mice.

Materials and methods: The tests were carried out on 35 adult male albino mice (16-20 weeks, weighed 30-35 g). Animals were categorized into seven groups each with five mice; the first group was treated with phosphate buffer saline (PBS) as negative control group. The second and third groups were treated daily and orally with two different doses of the DZN (60 and 90 mg/kg body weight) respectively dissolved in corn oil. Animals of fourth, fifth and sixth groups were orally administered grape seed oil at a dose of 2 g/kg body weight, after 6 hours the fourth and fifth groups subjected to DZN at the same dose given to second and third groups. Animals of the seventh group were supplied with corn oil. The treatments were continued for five weeks.

Results: The results showed that DZN increased the frequency of both chromatid and chromosome aberrations in bone marrow cells and sperm abnormalities. Treatment with grape seed oil extract showed a significant decrease ($P < 0.01$) on both total abnormal chromosomes and sperms, and there was no significant difference between these groups and negative control in most analyzed parameters. The dose of grape seed oil has significantly minimized the effect of first dose of DZN concerning chromosome structure while it worked in the second dose regarding sperm abnormalities.

Conclusions: This finding suggests the protective action of grape seed oil against DZN induced clastogenicity and sperm abnormalities.

Key words: chromosomal aberrations, sperm abnormalities, diazinon, grape seed oil.

Introduction:

The pollution of the environment plays a great role in the occurrence of many diseases affecting humans, animals, and plants. One of the major factors causing contamination of the environment is the wrong use of organophosphorus pesticides [1]. The contact with organophosphorus pesticides is a serious health problem for agricultural workers [2].

Diazinon (DZN) is a widespread used organophosphorus insecticide. It has been used since 1956 for the control of pests and soil insects, on fruits, and on flourish plants, vegetables and field crops [3]. Its residential uses have been revoked since 2004 by US Environmental Protection Agency, but its agricultural application is still very common [4]. It is used to control flies, greenhouses, animal facilities, places and other institu-

tions and public places where food or animal decays might be collected [5]. DZN can be highly toxic to human and animals [6, 7]. The mechanism of action of DZN is acetylcholinesterase enzyme inhibition [8], resulting in cholinergic overstimulation [9]. However, it may cause an imbalance in the free radicals production /elimination processes with the indirect reason of cellular damage [8, 10, 11, 12].

The plant products continuously play the basic role in treatments of about 80–85% of the world's population. Although the trends of chemistry and molecular biology providing fast growth of synthesized de novo drugs, plants remain a regular origin of medicinal compounds; up to 40% of new drugs may directly or indirectly be correlated to natural compounds [13].

Grape (*Vitis vinifera*) is one of the world's abundant fruit crops, and grape seed extract (GSE) is a complex matrix containing 16% oil, 11% proteins, 40% fiber, and 7% complex phenols including tannins, in addition to mineral salts and sugars [14]. It is a rich source of one of the most beneficial groups of plant flavonoids and proanthocyanidins oligomers [15]. GSE

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contains mainly flavonoids, which involved in reduction the oxidative stress in vitro and in vivo [16].

[17] Stated that GSE considered as a powerful antioxidant nutritive supplement that prevents premature aging and diseases. Oil of grape seeds regarded as a high potent antioxidant because of its rich source of polyphenolics [18].

[18, 19] showed that extracts obtained from grape seed have antimutagenic and anticarcinogenic effects by inhibiting enzymes of free radicals productions. The consumption of grape seed extracts may be useful in decreasing the side effects of chemotherapeutic agents in cancer treatment [20].

The present study was aimed to investigate the protective role of grape seed oil supplementation on chromosomal aberrations and sperm abnormalities induced by DZN toxicity in male mice.

Materials and methods:

The present study was carried out at Salahaddin University/ college of science/ Department of biology. The grape seed extracted oil was provided by Basso Fedele and Figli srl company (Italy).The experiments were carried out on 35 adult male albino mice (16-20 weeks, weighed 30-35 g), and were kept under constant environmental conditions with a 12:12 light-dark cycle. The animals were provided standard granulated chow. In the present work, animals were arranged into seven groups each with five mice; the first group was treated daily and orally with phosphate buffer saline (PBS) as negative control group. The second and third groups were treated daily and orally with two different doses of the DZN (60, 90 mg/kg body weight) in corn oil for five weeks. Animals of fourth and fifth groups were orally administered grape seed oil at a dose of 2 g/kg body weight (0.1 ml/animal/day) and after 6 hours subjected to DZN at the same dose given to second and third groups, daily for five weeks. Animals of the sixth group were treated with grape seed oil at the same dose given to groups 4 and 5. Animals of the

seventh group were supplemented with 0.1 ml of corn oil.

Then at the time of data harvesting chromosomal preparations were done by the standard method of [21]. Sperm was taken from epididymis using the method of [22] and [23].

Statistical analysis:

All data are expressed as means \pm standard error (M \pm SE) and statistical analysis were carried out using SPSS version 21. Comparisons between groups were made using one-way analysis of variance (ANOVA) with performing Duncan t-test as post hoc analysis. P values \leq 0.05 were considered as significant difference.

Results:

Table (1) summarizes the treatment effect of grape seed oil doses against DZN induced chromosomal aberrations.

A significant induction ($p < 0.01$) was found in two different doses of DZN on the total abnormal chromosome. Most types of aberrations were increased when compared with negative control group, while the acentric fragment was the only abnormality recorded there was non-significant effect in the second dose of DZN (90 mg/kg/day) (Figure 1).

The centromeric gap (8.60 \pm 0.24) in second dose of DZN (90 mg/kg/day) and (4.80 \pm 0.37) in first dose of DZN (60 mg/kg/day) showed the highest values of aberrant types respectively, while polyploidy (1.40 \pm 0.24) in the first dose and acentric fragment (1.00 \pm 0.00) in the second dose recorded the lowest values.

Treatment with grape seed oil extract showed significant protection ($P < 0.01$) for total abnormal metaphase including most studied aberrations when compared with positive control. The frequency of aberrations was not significant with negative control. It was apparent from the Table (1) that the dose of grape seed oil was have minimized the effect of DZN on chromosome structure, the most protective effects of grape seed oil was worked with the first dose of DZN.

Table1. Effects of grape seed oil extract against diazinon induced chromosomal aberrations in male albino mice. (Mean \pm SE) ($P < 0.01$).

	Total normal Chromosome	Total abnormal Chromosome	Centromeric gap	Centromeric break	Chromatid gap	Chromatid break	Ring chromosome	Dicentric chromosome	Pulverization	Polyploidy	Acentric fragment
Control	89.80 \pm 0.58 ^d	10.20 \pm 0.58 ^a	2.60 \pm 0.40 ^a	1.60 \pm 0.24 ^a	1.20 \pm 0.20 ^a	0.40 \pm 0.24 ^a	1.60 \pm 0.24 ^a	0.40 \pm 0.40 ^a	1.00 \pm 0.31 ^a	0.20 \pm 0.20 ^a	1.20 \pm 0.20 ^a
DZN 60 mg/Kg/day	77.40 \pm 0.92 ^b	22.60 \pm 0.92 ^c	4.80 \pm 0.37 ^b	2.60 \pm 0.24 ^{ab}	2.40 \pm 0.40 ^{bc}	2.40 \pm 0.24 ^b	2.80 \pm 0.20 ^b	2.00 \pm 0.00 ^b	2.00 \pm 0.31 ^{ab}	1.40 \pm 0.24 ^{bc}	2.20 \pm 0.37 ^b
DZN 90 mg/Kg/day	74.20 \pm 1.42 ^a	25.80 \pm 1.42 ^d	8.60 \pm 0.24 ^c	3.00 \pm 0.63 ^b	2.60 \pm 0.24 ^c	2.20 \pm 0.58 ^b	2.60 \pm 0.24 ^b	1.80 \pm 0.37 ^b	2.00 \pm 0.31 ^{ab}	2.00 \pm 0.44 ^c	1.00 \pm 0.00 ^a
DZN 60 mg/Kg/day + Grape oil (2g/kg/day)	82.00 \pm 1.30 ^c	18.00 \pm 1.30 ^b	3.00 \pm 0.44 ^a	2.00 \pm 0.31 ^{ab}	1.60 \pm 0.24 ^{ab}	0.80 \pm 0.20 ^a	2.80 \pm 0.20 ^b	2.00 \pm 0.00 ^b	2.20 \pm 0.20 ^b	1.60 \pm 0.24 ^{bc}	2.00 \pm 0.54 ^{ab}

DZN 90 mg/ Kg/day + Grape oil (2g/ kg/day)	79.80 ± 1.39 ^{bc}	20.20 ± 1.39 ^{bc}	3.60 ± 0.24 ^a	2.60 ± 0.24 ^{ab}	3.00 ± 0.54 ^c	2.40 ± ±0.24 ^b	3.00 ± 0.54 ^b	1.60 ± 0.67 ^b	1.40 ± 0.24 ^{ab}	0.80 ± 0.37 ^{ab}	1.80 ± 0.37 ^{ab}
Grape oil (2g/ kg/day)	89.60 ± 0.50 ^d	10.40 ± 0.50 ^a	2.80 ± 0.20 ^a	2.00 ± 0.31 ^{ab}	1.20 ± 0.20 ^a	0.00 ± ±0.00 ^a	1.60 ± 0.40 ^a	0.20 ± 0.20 ^a	1.20 ± 0.37 ^{ab}	0.20 ± 0.20 ^a	1.20 ± 0.20 ^a
Corn oil (2g/ kg/day)	89.00 ± 0.54 ^d	11.00 ± 0.54 ^a	3.40 ± 0.24 ^a	1.80 ± 0.20 ^a	1.20 ± 0.20 ^a	0.40 ± ±0.24 ^a	1.40 ± 0.24 ^a	0.40 ± 0.40 ^a	1.20 ± 0.37 ^{ab}	0.20 ± 0.20 ^a	1.00 ± 0.00 ^a

Note: Similar letters in each column refer to non significant difference while different letters refer to significant difference between them.

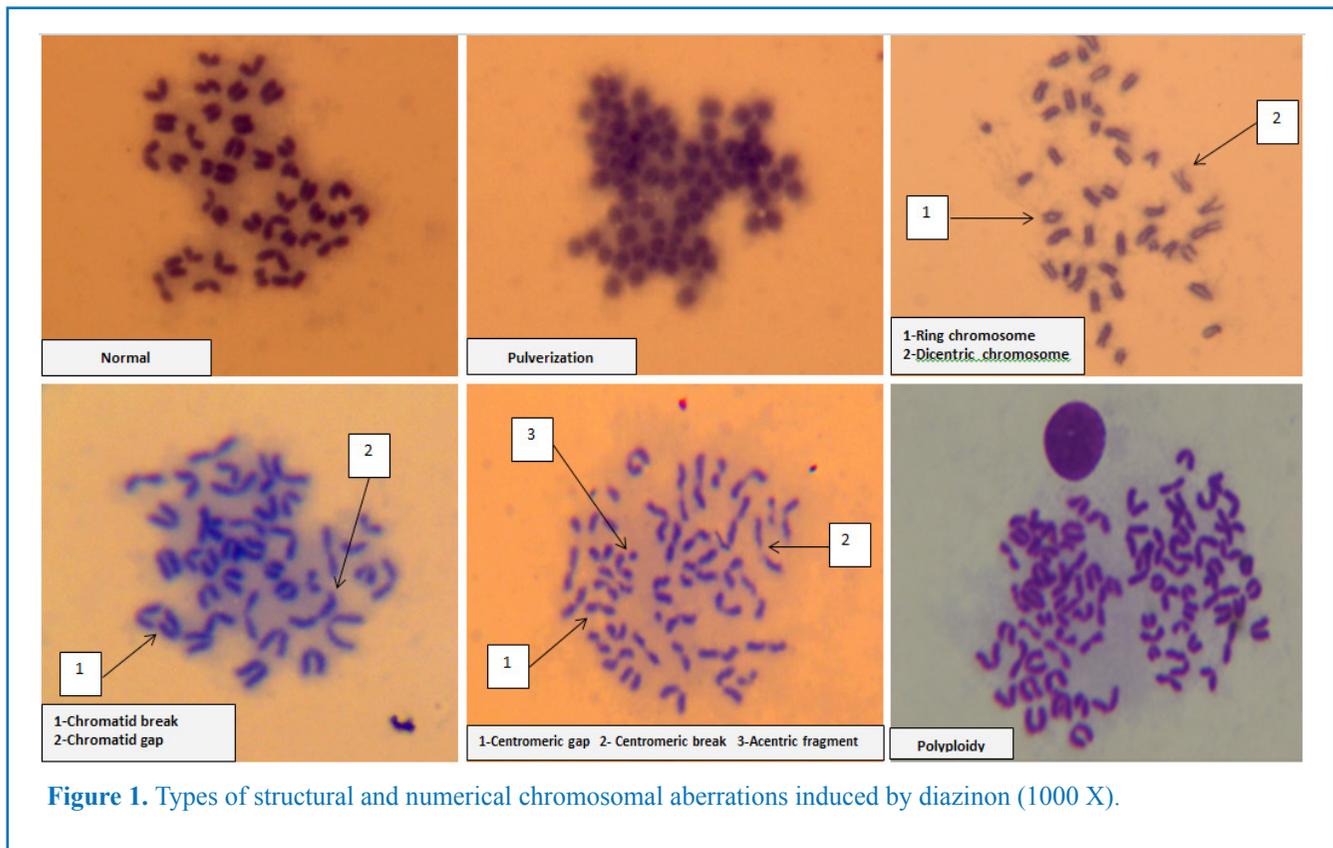


Figure 1. Types of structural and numerical chromosomal aberrations induced by diazinon (1000 X).

Table (2) showed the effect of grape seed oil treatment against DZN induced sperm abnormalities. Highly significant increase at ($P < 0.01$) were found in both DZN-treated groups (positive control) regarding the total abnormal sperms including (sperm without head, sperm without tail, sperm without hook, swollen head sperm, defective head sperm, and blunt hook sperm) when compared with negative control (PBS), but double head sperm was showed non-significant difference ($P < 0.01$) in the first treated group (Figure 2).

The highest value of abnormality type was sperm without tail (10.80 ± 1.24), (7.80 ± 0.96) in the first and second dose of DZN respectively, while the lowest value was double tail sperm (0.40 ± 0.24) in the first dose and double head sperm (2.40 ± 0.50) in the second dose respectively. Treatment with grape seed oil extract showed the significant decrease ($P < 0.01$) in total abnormal sperm including all studied sperm abnormalities compared

to the positive control, while there was no significant difference between grape treated groups and negative control in most analyzed parameters. From the Table (2) it is clear that the dose of grape seed extract decreased the effect of DZN on sperm morphology, the most protective effects of grape seed oil were worked with the second dose of DZN.

Table2. Effects of grape seed oil extract against diazinon induced sperm abnormalities in male albino mice. (Mean \pm SE) (P<0.01).

	Total normal sperm	Total abnormal sperm	Sperm without tail	Sperm without head	Sperm without hook	Double head sperm	Double tail sperm	Swollen head sperm	Defective head sperm	Blunt hook sperm
Control	\pm 89.20 e 0.73	\pm 10.80 a 0.73	0.40 ± 2.60 a	0.50 ± 2.40 ab	\pm 1.80 a 0.20	\pm 0.20 a 0.20	\pm 0.00 a 0.00	\pm 1.60 a 0.24	0.20 ± 1.20 a	0.00 ± 1.00 ab
DZN 60 mg/ Kg/day	\pm 63.80 a 2.03	\pm 36.20 e 2.03	\pm 10.80 e 1.24	1.06 ± 7.20 c	0.81 ± 3.60 b	\pm 1.00 a 0.31	0.24 ± 0.40 ab	\pm 4.00 b 0.31	0.37 ± 4.80 c	0.40 ± 4.40 d
DZN 90 mg/ Kg/day	\pm 68.40 b 1.02	\pm 31.60 d 1.02	0.96 ± 7.80 d	0.37 ± 4.20 b	0.37 ± 3.80 b	\pm 2.40 b 0.50	\pm 2.60 d 0.40	\pm 4.60 b 0.60	0.40 ± 3.60 bc	0.4 ± 2.60 c
DZN 60 mg/ Kg/day + Grape oil (2g/ (kg/day)	\pm 80.40 d 1.20	\pm 19.60 b 1.20	0.54 ± 5.00 bc	0.37 ± 3.80 ab	0.24 ± 2.60 ab	\pm 1.40 ab 0.74	0.58 ± 1.20 bc	\pm 2.40 a 0.60	0.24 ± 2.40 ab	0.20 ± 0.80 a
DZN 90 mg/ Kg/day + Grape oil (2g/ (kg/day)	\pm 73.60 c 1.40	\pm 26.40 c 1.43	0.24 ± 5.60 c	0.31 ± 3.00 b	0.40 ± 3.60 b	\pm 2.40 b 0.24	0.20 ± 1.80 cd	\pm 4.40 b 0.67	0.67 ± 3.60 bc	0.70 ± 2.00 bc
Grape oil (2g/ (kg/day)	\pm 89.60 e 0.50	\pm 10.40 a 0.50	0.24 ± 2.60 a	0.54 ± 2.00 a	\pm 1.40 a 0.24	\pm 0.20 a 0.20	\pm 0.00 a 0.00	\pm 1.40 a 0.24	0.24 ± 1.60 a	0.20 ± 1.20 ab
Corn oil (2g/ (kg/day)	\pm 86.60 e 1.63	\pm 13.40 a 1.63	0.58 ± 3.20 ab	0.60 ± 2.60 ab	\pm 2.20 a 0.20	\pm 0.20 a 0.20	\pm 0.00 a 0.00	\pm 1.80 a 0.20	0.44 ± 2.00 a	0.24 ± 1.40 ab

Note: Similar letters in each column refer to non significant difference while different letters refer to significant difference between them.

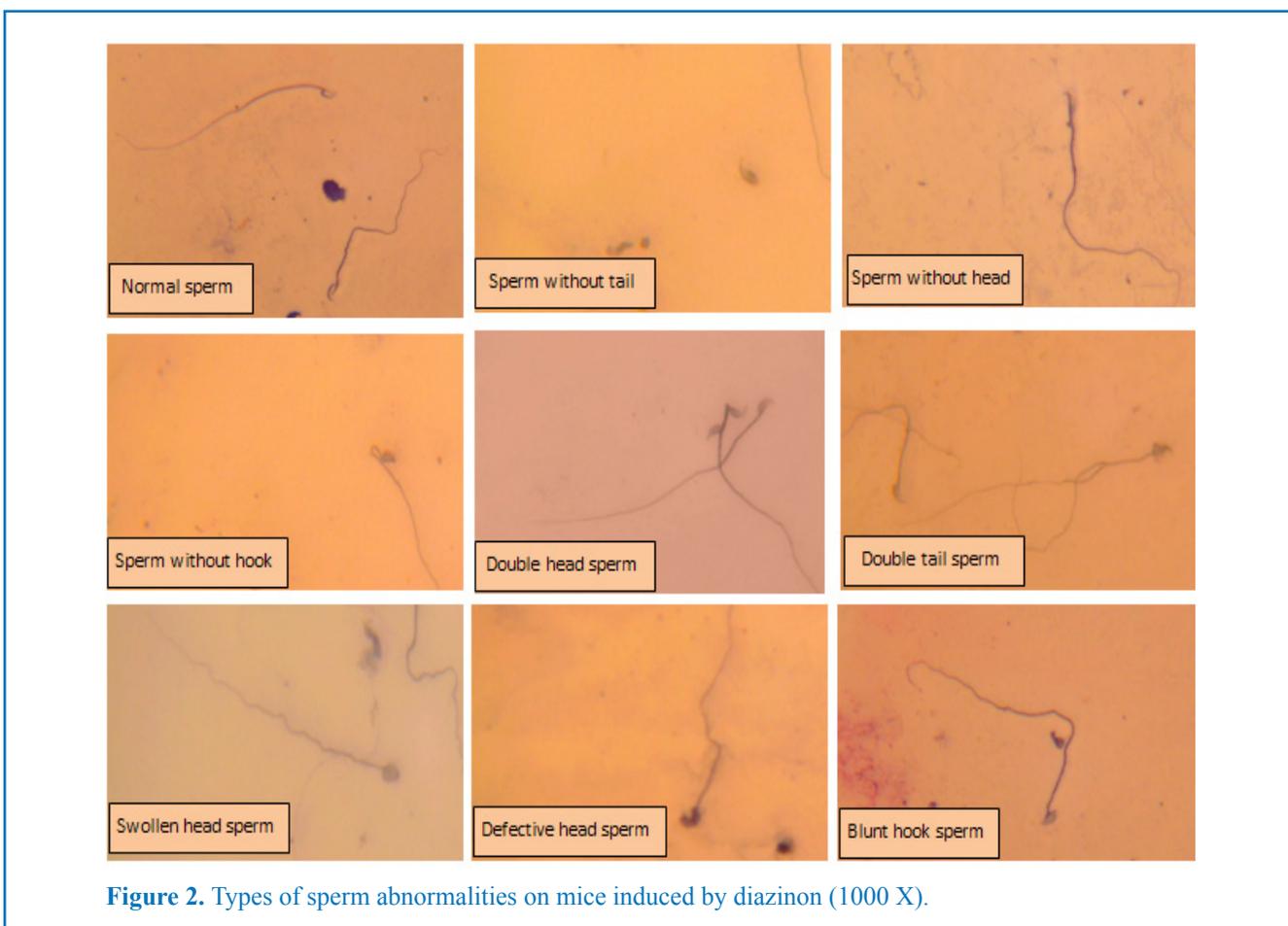


Figure 2. Types of sperm abnormalities on mice induced by diazinon (1000 X).

Discussion:

The undesired effects of pesticides have been identified as a serious public health concern during the past decades. [3].

The present study was conducted to examine the possible protective activity of orally administered grape seed oil against DZN induced cytotoxicity and genotoxicity towards mice somatic cells and sperm morphology *in vivo*.

Pretreatment of mice with grape seed oil for five weeks and simultaneously with doses of DZN significantly decreased the frequency of chromosomal aberrations in bone marrow cells and sperm abnormalities. These results agree with Abd El-Rahim and Hafiz (2009) [24] whom evaluated the protection conferred by grape seed oil and linseed oil against cyclophosphamide-induced bone marrow chromosomal aberrations and sperm abnormalities in adult Swiss albino mice. Who concluded that each of grape seed and linseed oils serves as an excellent antioxidant that limited genotoxicity of bone marrow cells and sperm abnormalities. Grape seed oil and wheat germ oil supplementation significantly decrease the toxic effects of chlorpyrifos-induced oxidative stress and caused a significant change in different biochemical parameters in the liver of male albino rats [25]. The administration of grape seed oil before gamma-radiation exposure may be a promising attempt in attenuating the extent of oxidative damage accompanying radiotherapy [26].

Additionally, [27] concluded that the grape seed oil plays a significant role in decreasing the effects of acrylamide intoxication in male rat genital organs especially in testes, epididymis, prostate glands and seminal vesicles that lead to infertility.

The exposure to pesticides including DZN produced testicular damage, which resulted in the spermatogenic arrest [3, 28, 29, 30].

Diazinon (DZN) caused a significant decrease in sperm counts and spermatogenic, Leydig and Sertoli cells and a reduction in serum testosterone concentration. Also, a significant decrease was recognized in diameter and weight of testes after DZN administration [31].

[32] Suggested that the direct effects of the DZN induced severe necrosis in the germinal cells and remarkable germinal cells degeneration lowered the sperm quality and quantity. Therefore the high content of pressured germinal cells, elevated abnormal, immature, death sperms and high infiltration of immune cells.

Grape seed extract acts as a potent antioxidant prevented genotoxicity of bone marrow cells by reducing a total number of aberrant cells and different types of structural chromosomal aberrations caused by mutagen [33]. Also, [34] reported that grape seed inhibits the mutagenic effects of cyclophosphamide and/or other mutagens in rats and mice, both *in vitro* and/or *in vivo*.

Grape seed extracts prevented DNA oxidative damage in various tissues and DNA fragmentation induced by many agents [33, 35, 36].

[37] Proposed that *in vivo* protection of DNA by grape seed

extracts might be due to detoxification of cytotoxic radicals and considered the contribution to DNA repair. The cause of the antigenotoxic effect of the grape seed extracts is the presence of a lot of biologically active compounds in it, mainly antioxidants. From a cellular perspective, one of the most advantageous features of proanthocyanidins oligomers free radical scavenging activity is chemical structure; it is incorporated into cell membranes. This physical characteristic along with its ability to protect against both water and fat-soluble free radicals provides unbelievable protection to the cells against free radical alterations on chromosomes [38].

Many researchers found the straight relations between genotoxicity and chromosomal instability induced by many agents with the parameters of oxidative stress [39, 40].

The elevated oxidative stress due to treatment with cisplatin might be the cause of chromosomal aberration and sperm shape abnormalities. Extract mixtures prepared from a polar and non-polar fraction of red grape, coriander, roselle, and fenel were shown to be efficient in lowering chromosomal aberration and abnormal sperm induced by cisplatin [41].

The presence of antioxidants and anti-inflammatory bioactive constituent in these extract mixtures might be the cause of reduction of such chromosomal aberration and sperm shape abnormalities. Stability of genome might be afforded by treatment with antioxidant, anti-mutagenic, anti-carcinogenic and anti-inflammatory bioactive constituent such as phenolic compounds [42]. Grape seed extract (GSE) significantly protected mice bone marrow chromosomes from gentamicin-induced genotoxicity by reducing different types of structural chromosomal aberrations and the total number of aberrant cells. So, the grape extracts mixture, acted as a potent antioxidant preventing kidney damage and genotoxicity of bone marrow cells, which agreed with a study of [33].

GSE significantly preserved mice bone marrow chromosomes from doxorubicin-induced genotoxicity by reducing the frequency of structural chromosomal aberrations and the total aberrant metaphases [43].

Pretreatment of mice with GSE significantly elevated the level of sperm motility reduced by the low dose of cisplatin. At the higher dose of cisplatin, the effect of GSE was not statistically significant comparing with the cisplatin group alone. The cause of the antigenotoxic effect of the GSE is the presence of a lot of biologically active compounds in it, mainly antioxidants that protect DNA through detoxification of free radicals. Many vegetables and fruits are known to prevent chromosomally and DNA damage in animals [44, 45]. Also [46] demonstrated that grape seed proanthocyanidins (GSPs) could improve functional activation of the immune system, and the antitumor effects of GSPs were achieved by immunostimulating properties.

Conclusion and recommendation:

The antioxidative effects of grape seed extracted oil may play an important role in cell protection from genotoxic effects of DZN. Considering the present results, it can be concluded that this study shows that the grape seed extracted oil is beneficial in lowering the clastogenicity and sperm abnor-

malities induced by DZN exposure in mice. Finally, further investigations are needed to explore the mechanism action of

grape seed extracted oil against DZN toxicity.

References:

1. Al-Haj, M.; Nasser, A. and Anis, A. (2005). Survey of pesticides used in Qat cultivation in Dhale and Yafe and their adverse effects. *J. Nat. Appl. Sci.* 9: 103–110.
2. Hurtig, A.K. San Sebastian, M.; Soto, A.; Shingre, A.; Zambrano, D. and Guerrero, W. (2003). Pesticide use among farmers in the Amazon basin of Ecuador. *Arch. Environ. Health.* 58: 223–228.
3. Al-Attar, A.M. (2015). Effect of grapeseed oil on diazinon-induced physiological and histopathological alterations in rats Saudi Journal of Biological Sciences. 22: 284–292.
4. Edwards, D. (2006). Reregistration eligibility decision for diazinon. In: Finalization of Interim Reregistration Eligibility Decisions (IREDs). Available via: US Environmental Protection Agency Office of Pesticide Programs, US Environmental Protection Agency Washington, DC. http://www.epa.gov/oppsrrd1/REDs/diazinon_red.pdf. Accessed 27 Nov 2012.
5. Dikshith, T.S.S. and Diwan PV. (2003). Industrial Guide to Chemical and Drug Safety. Wiley-IEEE, 189–191.
6. Poet, T.S.; Kousba, A.A.; Dennison, S.L. and Timchalk, C. (2004). Physiologically base pharmacokinetic/pharmacodynamic model for the organophosphorus pesticide diazinon. *Neurotoxicology.* 25: 1013–1030.
7. Sarabia, L.; Maurer, I. and Bustos-Obregón, E. (2009). Melatonin prevents damage elicited by the organophosphorous pesticide diazinon on the mouse testis. *Ecotoxicol. Environ. Saf.* 72: 938–942.
8. Kamanyire, R. and Karalliedde, L. (2004). Organophosphate toxicity and occupational exposure. *Occup. Med.* 54: 69–75.
9. International Programme on Chemical Safety. (1998). In: United Nations Environment Programme, International Labour Organisation, and World Health Organization, editor. Diazinon (environmental health criteria; 198). Geneva: WHO; 1998.
10. Gokcimen, A.; Gulle, K.; Demirin, H.; Bayram, D.; Kocak, A. and Altuntas, I. (2007). Effects of diazinon at different doses on rat liver and pancreas tissues. *Pestic. Biochem. Physiol.* 87: 103–108.
11. Roegge, C.S.; Timofeeva, O.A.; Seidler, F.J.; Slotkin, T.A. and Levin, E.D. (2008). Developmental diazinon neurotoxicity in rats: later effects on emotional response. *Brain Res. Bull.* 75: 166–172.
12. Cakici, O. and Akat, E. (2013). Effects of oral exposure to diazinon on mice liver and kidney tissues: biometric analyses of histopathologic changes. *Anal. Quant. Cytol. Histol.* 35: 7–16.
13. Solyanik, G.I.; Fedorchuk, A.G.; Pyaskovskaya, O.N.; Dasyukevitch, O.I.; Khranovskaya, N.N.; Aksenov, G.N. and Sobetsky, V.V. (2004). Anticancer activity of acetonitrile-containing herbal extract BC1. *Exp. Oncol.* 26: 307–311.
14. Shi, J.; Jianmel, Y.; Joseph, E.P. and Yukio, K. (2003). Polyphenolics in grape seeds biochemistry and functionality. *J. Med. Food.* 6: 291–299.
15. El-Ashmawy, I.M.; Saleh, A. and Salama, O.M. (2007). Effects of marjoram volatile oil and grape seed extract on ethanol toxicity in male rats. *Basic & Clinical Pharmacology & Toxicology.* 101: 320–327.
16. Martinez-Florez, S.; Gonzalez-Gallego, J. and Culebras, J.M. (2002). Flavonoids: properties and antioxidizing action. *Nutr. Hosp.*, 17: 271–278.
17. Chedea, V.S.; Braicu, C. and Socaciu, C. (2010). Antioxidant / prooxidant activity of polyphenolic grape seed extract. *Food Chem.*, 121: 132–139.
18. Maier, T.; Schieber, A.; Kammerer, D. and Carle, R. (2009). Residues of grape (*Vitis vinifera* L.) seed oil production as a valuable source of phenolic antioxidants. *Food Chemistry.* 112: 551–559.
19. Li, W.G.; Zhang, X.Y. and Wu, Y.J. (2001). Anti-inflammatory effect and mechanism of proanthocyanidins from grape seeds. *Acta Pharmacol. Sin.*, 22: 1117–1120.
20. Aysun, C.; Leylagul, K.; Ismail, K.; Sibel, H.; Recep, S.; Ahmet, O.; Okan, O. and Osman, S. (2008). The effect of grape seed extract on radiation-induced oxidative stress in the rat liver. *Turk J. Gastroenterol.*, 19 (2): 92–98.
21. Evans, E.P.; Breckon, G. and Ford, C. F. (1964). An air drying method for meiotic preparation from mammalian testis *Cytogenetic.* 41(6):317–321. (Cited by Al-Attar, M.S.M. (2004). Evaluation of pesticides toxicity on laboratory mice and human chromosome in Iraqi-Kurdistan. Ph.D. Thesis. College of Education, University of Salahaddin- Erbil, Iraq.
22. Karanowska, L. (1976). Chromosomal mutation. (Cited by R. Sahai, *Advances in cytogenetics and their application in livestock improvement and production*, N.D.R.I. Karnal, India, (35): 136).
23. Wyrobek, A.J. and Bruce, W.R. (1975). Chemical induction of sperm abnormalities in mice. *Proc. Nat. Acad. Sci. USA.* 72(11):4425–4429.
24. Abd El-Rahim, A.H. and Hafiz, N.A. (2009). Investigation on the protective effect of grape seed and linseed oils against cyclophosphamide induced genotoxicity in mice. *Global Vet.* 3: 377–382.
25. Khalifa, F.K.; Khalil, F.A.; Barakat, H.A. and Hassan, M.M. (2011). Protective role of wheat germ and grape seed oils in chlorpyrifos-induced oxidative stress, biochemical and histological alterations in liver of rats. *Aust. J. Basic Appl. Sci.* 5: 54–66.
26. Naguib, N.I. (2011). Grape seed oil extract protects against radiation-induced oxidative damage in rat's eye. *Isotope Rad. Res.* 43: 1255–1264.
27. Hasseeb, M.M.; Al-Hizab, F.A. and Hamouda, M.A. (2013). Impacts of grape seed oil supplementation against the acrylamide induced lesions in male genital organs of rats. *Pak. Vet. J.* 33: 282–286.
28. Adamkovicova, M.; Toman, R.; and Cabaj, M. (2010). Diazinon and cadmium acute testicular toxicity in rats examined by histopathological and morphometric methods. *Slovak J. Anim. Sci.* 43: 134–140.
29. Jorsaraei, S.G.A.; Firoozjaee, A.; Pasha, Y.Y.; Marzony, E.T. and Sarabi, E. (2010). Histopathological effects of single dose treatment of diazinon on testes structure in rat. *Yakhteh Med. J.* 12: 39–42.
30. Zari, T.A. and Al-Attar, A.M. (2011). Therapeutic effects of olive leaves extract on rats treated with a sublethal concentration of carbendazim. *Eur. Rev. Med. Pharmacol. Sci.* 15, 413–426.
31. Fattahi, E.; Parivar, K.; Jorsaraei, S.G.A. and Moghadamnia, A.A. (2009). The effects of diazinon on testosterone, FSH and LH levels and testicular tissue in mice. *Iran. J. Reprod. Med.* 7: 59–64.
32. Ali, K.; Reza, N.G.; Syamak, S.; Aref, H.; Ehsan, H.; Leila, R.; Mohammad, B.; Ali, N.; Davoud, K. and Esmaiel, G. (2011). Protective effect of selenium on diazinon induced determination impact on the testes in mature male rats. *Global Vet.* 7: 370–380.
33. El-Ashmawy, I.M.; El-Nahas, A.F. and Salama, O.M. (2006). Grape seed extract prevents gentamicin-induced nephrotoxicity and genotoxicity in bone marrow cells of mice. *Basic Clin. Pharmacol. Toxicol.*, 99: 230–236.
34. Asita, A.O.; Dingann, M.E. and Magama, S. (2008). Lack of modu-

- latory effect of asparagus, tomato and grape juice on cyclophosphamide-induced genotoxicity in mice. *African J. Biotechnol.*, 7: 3383-3388.
35. Llopiz, N.; Puiggros, F.; Cespedes, E.; Arola, L.; Ardevol, A.; Blade, C. and Salvado, M.J. (2004). Antigenotoxic effect of grape seed procyanidin extract in Fao cells submitted to oxidative stress. *J. Agric. Food Chem.*, 52: 1083-1087.
 36. Fan, P.H. and Lou, H.X. (2004). Isolation and structure identification of grape seed polyphenols and its effects on oxidative damage to cellular DNA. *Yao Xue Xue Bao.*, 39: 869-875.
 37. Ray, S.D.; Patel, D.; Wong, V. and Bagchi, D. (2000). In vivo protection of DNA damage associated apoptotic and necrotic cell deaths during acetaminophen-induced nephrotoxicity, amiodarone-induced lung toxicity and doxorubicin-induced cardiotoxicity by a novel IH636 grape seed proanthocyanidin extract. *Res. Commun. Molecular Pathol. Pharmacol.*, 107:137-166.
 38. Bagchi, D.; Ray, S.D.; Patel, D. and Bagchi, M. (2001). Protection against drug-and chemical-induced multiorgan toxicity by a novel IH636 grape seed proanthocyanidin extract. *Drugs Experimental. Clinical Res.*, 27:3-15.
 39. Jahangir, T.; Khan, T.H.; Prasad, L. and Sultana, S. (2005). Reversal of cadmium chloride induced oxidative stress and genotoxicity in Swiss albino mice. *J. Pharmacy and Pharmacol.*, 57: 1199-1204.
 40. Lee, D.H.; Lim, B.S.; Lee, Y.K.; Ahn, S.J. and Yang, H.C. (2006). Involvement of oxidative stress in mutagenicity and apoptosis caused by dental resin monomers in cell cultures. *Dent. Mater.*, 22: 1086-1092.
 41. Al-Okbi, S.Y.; Mohamed, D. A.; Hamed, TH. E.; Esmail, R. SH. and Donya, S.M. (2014). Plant Food Extracts as a Source of Bioactive Compounds for Prevention of Cisplatin-Induced Kidney Dysfunction in Rats. *Pol. J. Food Nutr. Sci.*, 64 (1): 49-57.
 42. Ferguson, L.R. (2001). Role of plant polyphenols in genomic stability. *Mut. Res.*, SI, 475: 89-111.
 43. Yalçın, E.; Oruç, E.; Cavuşoğlu, K. and Yapar, K. (2010). Protective role of grape seed extract against doxorubicin-induced cardiotoxicity and genotoxicity in albino mice. *J. Med. Food.* 13: 917-25.
 44. Nersesyan, A. and Muradyan, R. (2004). Sea-buckthorn juice protects mice against genotoxic action of cisplatin. *Exp. Oncol.* 26:153-5.
 45. Miyata, M.; Takano, H.; Guo, L.Q.; Nagata, K. and Yamazoe, Y. (2004). Grapefruit juice intake does not enhance but rather protects against aflatoxin B1-induced liver DNA damage through a reduction in hepatic CYP3A activity. *Carcinogenesis.* 25:203-9.
 46. Tong, H.; Song, X.; Sun, X.; Sun, G. and Du, F. (2011). Immunomodulatory and antitumor activities of grape seed proanthocyanidins. *Journal of Agricultural & Food Chemistry.* 59(21):11543-11547.

التأثيرات المضادة للتطهير لمستخلص زيت بذور العنب على مبيد الديازينون في الفئران المختبرية

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الخلاصة:

تعد المخاطر على صحة الإنسان بسبب التعرض للملوثات الكيميائية من المشاكل العالمية المزمنة. وتشكل المبيدات المواد الكيميائية السامة الهامة في البيئة. بحثت الدراسة الحالية تأثير مستخلص زيت بذور العنب على الديازينون (DZN) المسبب للتغيرات الكروموسومية وتشوهات الحيوانات المنوية في ذكور الفئران المختبرية. تم إجراء الاختبارات على 35 من ذكور الفئران المختبرية البيضاء (بعمر 16-20 أسبوعاً ووزن 30-35 غم). قسمت الحيوانات إلى سبعة مجاميع لكل منها خمسة فئران. أعطيت المجموعة الأولى المحلول المتعادل (PBS) كمجموعة مقارنة سلبية. وأعطيت المجموعتين الثانية والثالثة يومياً وعن طريق الفم الجرعتين من مبيد الديازينون 60 و 90 ملغم / كغم من وزن الجسم) المذاب في زيت الذرة على التوالي. الحيوانات في المجموعات الرابعة والخامسة والسادسة أعطيت وعن طريق الفم زيت بذور العنب بتركيز 2 غرام / كغم من وزن الجسم، وبعد 6 ساعات المجموعتين الرابعة والخامسة أعطيت DZN بنفس الجرعات المعطاه المجموعتين الثانية والثالثة. تم معاملة الحيوانات في المجموعة السابعة بزيت الذرة. واستمر التجربة لمدة خمسة أسابيع. وأظهرت النتائج أن المبيد DZN يعمل على زيادة وتيرة التغيرات الكروماتيدية والانحرافات الكروموسومية في خلايا نخاع العظام وتشوهات الحيوانات المنوية. وأظهرت المعاملة مع مستخلص زيت بذور العنب انخفاضاً معنوياً ($P < 0.01$) في كل من إجمالي الكروموسومات غير الطبيعية والحيوانات المنوية، وكان هناك فرق كبير بين هذه المجاميع ومجموعة المقارنة السلبية في معظم المعاملات المدروسة. إن جرعة زيت بذور العنب أدت إلى تقليل معنوي في تأثير الجرعة الأولى من المبيد على التغيرات الكروموسومية، بينما أدت جرعة زيت بذور العنب إلى تقليل معنوي في تأثير الجرعة الثانية للمبيد في تشوهات الحيوانات المنوية. وتقدر هذه النتيجة بأن زيت بذور العنب له تأثير وقائي ضد مبيد الديازينون DZN المسبب للتغيرات الكروموسومية وتشوهات الحيوانات المنوية في ذكور الفئران المختبرية.